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Infectious diseases of reptiles: An overview



African spurred tortoise (*Centrochelys sulcata*) with rhinitis

Photo source: PD Dr. Rachel Marschang

In recent years, our understanding of infectious diseases in reptiles has grown immensely. It is also clear that the connection between infection and disease is often dependent on multiple factors, including pathogen specific factors (e.g. strain specific properties, virulence factors) and host specific factors (e.g. species, age, sex), as well as environmental conditions (e.g. temperature, hygiene, time of year) and coinfections with other agents.

Infections in reptiles also often persist, making quarantine particularly important in this group of animals. The diagnosis of infectious agents in reptiles has also made great strides in recent years. The following pages contain an overview of select infectious agents found in tortoises and turtles, snakes, and lizards. The overview is mainly focused on microorganisms (viruses, bacteria, and fungi), although some parasites are also included. If you are unsure what tests or what types of samples are best in a specific case, we are happy to consult with you on cases.

Select pathogens according to affected organ systems:

Tortoises and turtles

Skin: Herpes-, rana-, papillomaviruses, various bacteria, various fungi (USA: *Emydoimyces testavorans*); **Respiratory tract:**

Herpes-, picorna-, adeno-, rana-, paramyxoviruses (a.k.a. ferlaviruses), mycoplasma, chlamydia, other bacteria, fungi, intranuclear coccidia (TINC);

Gastrointestinal tract: Herpes-, adeno-, rana-, reoviruses, various bacteria, fungi, intranuclear coccidia (TINC), cryptosporidia, various other parasites

Snakes

Skin: Arena-, papillomaviruses, various bacteria, *Ophidiomyces ophidiicola*, other fungi; **Respiratory tract:** Nido-, arena-, adeno-, reo-, paramyxoviruses (a.k.a. ferlaviruses), sunshinevirus, mycoplasma, chlamydia, other bacteria, fungi, parasites;

Gastrointestinal tract: Adeno-, arena-, herpes-, rana-, reoviruses, various bacteria, fungi, cryptosporidia, other parasites; **CNS:** Arena-, paramyxoviruses (a.k.a. ferlaviruses), sunshinevirus, *Entamoeba invadens*

Lizards

Skin: Rana-, irido-, herpes-, reo, papilloma-, poxviruses, *Devriesea agamarum*, other bacteria, *Nannizziopsis* spp., other fungi;

Respiratory tract: Paramyxoviruses (a.k.a. ferlaviruses), adeno-, reoviruses, chlamdia, other bacteria and fungi; **Gastrointestinal tract:** Adeno-, herpes-, irido-, reoviruses, various bacteria, fungi, cryptosporidia, other parasites; **CNS:** Adenoviruses

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Table 1: Select infectious agents found in tortoises and turtles and their laboratory diagnosis

Infectious agent	Affected species	Affected tissues and clinical signs	Samples for diagnosis*	Methods
Adenoviruses	Tort and tort	From inapparent carriers to systemic disease and sudden death	Cloacal swabs, intestine, liver, other affected tissues	PCR
Herpesviruses	Various	Mostly upper respiratory and GI tract, skin, in some cases inapparent infection	Oral swab, cloacal swab, skin, tissues (tongue, liver, brain, intestine, others)	PCR Sero: VN in tort
Paramyxoviruses (ferlaviruses)	Esp. tort	Mostly pneumonia	Tracheal wash, oral and cloacal swabs, lung, other tissues	PCR
Picornavirus (virus „X“)	Tort	Softening of the carapace in juveniles, renal disease, rhinitis, in some cases inapparent infection	Oral swabs, cloacal swabs, various tissues	PCR, VI Sero: VN
Ranaviruses	Tort and tort	Upper respiratory and GI tract, liver, blood vessels	Oral and cloacal swabs often not sensitive, blood can be tested, tissue samples best for virus detection	PCR, VI
Reoviruses	Esp. tort	Respiratory tract, possibly also GIT	Oral and cloacal swabs, tissues	PCR, VI
Bacteria (aerob and anaerob)	All	Many facultative pathogens, can affect various tissues	Samples from lesions. Interpretation in conjunction with clinical signs	Culture
Chlamydia	Esp. tort	Granulomas, rhinitis, pneumonia, myocarditis, hepatitis	Nasal washes, oral and cloacal swabs, affected tissues	PCR
Mycobacteria	All	Esp. granulomas	Material from lesions	Histo, ZN
Mycoplasma	Tort and tort	URTD	Oral swabs, nasal washes	PCR
Fungi and yeasts	All	Many facultative pathogens, can affect various tissues	Samples from lesions. Interpretation in conjunction with clinical signs	Culture
Cryptosporidia	Esp. tort	Depending on the parasite species, the stomach or the intestine may be affected	Faeces, gastric or intestinal mucosa, possibly gastric lavage, cloacal swabs	PCR
Intranuclear coccidia (TINC)	Tort and tort	From inapparent carriers to systemic disease and sudden death	Cloacal swabs, oral swabs, faeces, tissues	PCR
Parasites	All	Esp. GIT, from inapparent carriers to severe infestations and death	Esp. faeces	N, flot

*The ideal sample depends on the stage of infection, type of pathogen, and host species and should be chosen based on the clinical question. Flot = flotation; GIT = gastrointestinal tract; Histo = histology; N = native; Sero = serology (antibody detection); tort = tortoise; turt = turtle; URTD = upper respiratory tract disease; VN = virus neutralisation test; ZN = Ziehl-Neelson stain

Table 2: Select infectious agents found in snakes and their laboratory diagnosis

Infectious agent	Affected species	Affected tissues and clinical signs	Samples for diagnosis*	Methods
Adenoviruses	All	Esp. GIT and liver	Cloacal swabs, faeces, intestine, liver	PCR, VI
Arenaviruses	Boas and pythons	Inclusion body disease (IBD)	Oesophageal swabs, blood, tissues (esp. brain, liver, kidney, lymph., pancreas)	PCR, cyto, histo
Herpesviruses	Various	Liver, kidney, oral mucosa, venom glands	Oral swabs, tissues	PCR
Nidoviruses	Esp. pythons	Lung, upper respiratory tract, other systems, in some cases inapparent infection	Oral swabs, cloacal swabs, tissues (lung)	PCR
Paramyxoviruses (ferlaviruses)	Various, esp. vipers, colubrids	Esp. respiratory tract	Tracheal wash, oral and cloacal swabs, lung	PCR Sero: HI
Ranaviruses	Various	Esp. liver, oral cavity	Oral and cloacal swabs often not sensitive, tissue samples best for testing	PCR, VI
Reoviruses	Various	Respiratory tract, GIT, CNS	Oral and cloacal swabs, tissues	PCR, VI
Sunshinevirus	Pythons	CNS, respiratory tract	Oral and cloacal swabs, tissues (esp. brain)	PCR
Bacteria (aerob and anaero)	All	Many facultative pathogens, can affect various tissues	Samples from lesions. Interpretation in conjunction with clinical signs	Culture
Chlamydia	Various	Granulomas, pneumonia, myocarditis, hepatitis, other tissues can also be affected	Oral and cloacal swabs, affected tissues	PCR
Mycobacteria	All	Esp. granulomas	Material from lesions	Histo, ZN
Mycoplasma	Esp. pythons	Upper respiratory tract	Oral swabs	PCR
Ophidiomyces ophiodiicola	Various	Skin	Skin (swabs, biopsies, shreds)	PCR
Fungi	Fungi and yeasts	All	Many facultative pathogens, can affect various tissues	Samples from lesions. Interpretation in conjunction with clinical signs
Cryptosporidia	Various	Stomach most often affected	Gastric lavage, regurgitated material, gastric mucosa	PCR, ZN, IFAT
Parasites (others)	All	Esp. GIT, inapparent carriers to severe infestations and death	Esp. faeces	N, float

*The ideal sample depends on the stage of infection, type of pathogen, and host species and should be chosen based on the clinical question. CNS = central nervous system; cyto = cytology; float = flotation; GIT = gastrointestinal tract; HI = haemagglutination inhibition; histo = histology; IFAT = immunofluorescence antibody test; lymph. = lymphatic tissue; N = native; sero = serology (antibody detection); VI = virus isolation in cell culture; ZN = Ziehl-Neelsen stain

Table 3: Select infectious agents found in lizards and their laboratory diagnosis

Infectious agent	Affected species	Affected tissues and clinical signs	Samples for diagnosis*	Methods
Viruses	Adenoviruses	All, esp. bearded dragons	Liver and GIT, in some cases CNS signs	CS, tissues (liver and intestine)
	Herpesviruses	Various	Liver, skin, oral mucosa	OS, CS, tissues
	Iridoviruses	Esp. bearded dragons, chameleons, others, also feed insects (crickets)	Skin, GIT	Tissues (not CS since virus can originate from feed insects)
	Nidoviruses	Shinglebacks, others	Upper respiratory tract	OS, tissues
	Paramyxoviruses (ferlaviruses)	Various	Esp. respiratory tract	Tracheal wash, OS, CS, tissues
	Ranaviruses	Various	Skin, liver, other tissues	Tissues samples; OS and CS not sensitive
	Reoviruses	Various	Respiratory tract, GIT	OS, CS, tissues
	Bacteria (aerob and anaerob)	All	Many facultative pathogens, can affect various tissues	Samples from lesions. Interpretation in conjunction with clinical signs
	Chlamydia	Various	Granulomas, pneumonia, myocarditis, hepatitis, others	OS, CS, affected tissues
	<i>Devriesea agamarum</i>	Esp. <i>Uromastyx</i> spp., others	Skin (cheilitis)	PCR
Bacteria	Mycobacteria	All	Esp. granulomas	Culture
	<i>Encephalitozoon pogonae</i>	Bearded dragons, other microsporidia described in other species	Liver, granulomas, others	Histo, ZN
	<i>Nannizziopsis</i> spp.	Various, esp. agamids	Skin, in some cases systemic disease	PCR
	Fungi and yeasts (others)	All	Many facultative pathogens, can affect various tissues	Culture
Parasites	Cryptosporidia	Various, common in leopard geckos	Esp. intestine	IFAT
	Parasites (others)	All	Esp. GIT, inapparent carriers to severe infestations and death	N, float
			Esp. faeces	

*The ideal sample depends on the stage of infection, type of pathogen, and host species and should be chosen based on the clinical question. CNS = central nervous system; CS = cloacal swab; float = flotation; GIT = gastrointestinal tract; HI = haemagglutination inhibition; histo = histology; IFAT = immunofluorescence antibody test; N = native; OS = oral swab; Sero = serology (antibody detection); VI = virus isolation in cell culture; ZN = Ziehl-Neelsen stain